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Hydroxyapatite and Biopolymer Composites with Promising Biomedical Applications

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ABSTRACT

The purpose of tissue engineering (regenerative medicine) is to develop materials that replace human tissue, having as main characteristics' biodegradability, biocompatibility, no toxicity, osteoconductivity, which lead to cell maturation and proliferation. Due to the importance of the development of this type of materials, several researchers have used biopolymers and calcium phosphate salts (hydroxyapatite) as composites to be used in this area as drug releases, scaffolds, implants, among others. Different biopolymers can be suitable for this type of application, in this work we have described the most widely used biopolymers for biomedical purposes, such as alginate, collagen, gellan gum, chitosan, and polylactic acid, in addition to a detailed description of hydroxyapatite, biopolymers, as well as biopolymer/hydroxyapatite composites, to highlight their potential and the most relevant characteristics of these materials.

KEYWORDS: Composites, Hydroxyapatite, Biopolymers

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INTRODUCTION

Currently, bone diseases are estimated to be one of the main causes of disability in humans. Osteoporosis, for example, causes approximately 9 million bone fractures worldwide with a high risk of morbidity and mortality of 12-20 % of sufferers ^[1]. A bone replacement must imitate the tissue by replicating its structural and functional requirements, in such a way that it generates an adequate response from the biological system to which it has been subjected ^[2]. To achieve this, the use of tissue engineering (TE) has been applied, which is a multidisciplinary science that uses the principles of engineering and life sciences to improve or develop biomaterials that generate a suitable microenvironment through the combination of cell ^[3]. The objective of TE is to produce biomaterials that are biocompatible, biodegradable, bioactive, capable of promoting cell proliferation, non-toxic, and osteoconductive, in order to provide similar characteristics to the cellular configuration of human tissue [4] [5].

Hence, several researches have developed biomaterials capable of satisfying this type of needs through the use of composite materials based on ceramics and biopolymers, since they have been shown to have a great similarity with bone tissue, since it consists of an inorganic phase (minerals such as calcium phosphate) and an organic phase (collagen and proteins) ^[6].

In this paper, given the importance of the development of this type of biomaterials, an extensive literature review has been carried out about the most utilized biopolymers in composites with inorganic hydroxyapatite phase, which have potential biomedical applications, describing the generalities of each of them, as well as recent research on this type of biomaterials.

MATERIALS AND METHODS

A bibliographic review was carried out to collect information about hydroxyapatite, biopolymers, as well as biopolymer/hydroxyapatite composites using the keywords hydroxyapatite, biopolymers, polymer matrix composites, tissue engineering, composite procurement, etc. Original papers and books were taken into account, both in English and Spanish, published in scientific journals indexed in Scopus, Science Direct, Google Scholar, and Web of Science. No limits on the year of publication were used, but the articles published in the last ten years were chosen for the description of how biopolymer/ hydroxyapatite composites are obtained, selecting a total of 132 references including both articles and books.

RESULTS AND DISCUSSION

Hydroxyapatite

The skeleton of the average adult is composed of 40% bone structure with size and shape defined for specific functions, the most important of which are to facilitate movement, protect vital organs, store minerals, deposit bone marrow, and provide the support structure of the body ^{[7] [8]}. Human bone is a complex dynamic tissue of dense nature, its extracellular matrix is composed mainly of collagen fiber (organic phase 30%) and the mineral part of which the major component is calcium phosphates (inorganic phase 70%). The inorganic content of the bone matrix is hydroxyapatite (Ha), which in combination with other biomaterials has been demonstrated to provide biocompatibility, biodegradability, and osteoconductivity ^{[4] [9]}.

Ha is closely related to basic calcium phosphate, which is the predominant inorganic material in the bones of all vertebrates, as well as in dental enamel. Has the chemical formula $Ca_{10}(PO_4)_6(OH)_2$, this phosphate crystallizes in a hexagonal form, see Figure 1, with a P63/m symmetry group, with lattice parameters a = b= 9.432 Å and c = 6.88 Å, it is commonly obtained in aqueous conditions and often tends to have particle sizes around 5 to 1 nm, so they have a large surface area, which intensifies its chemical activity. The ionic character of the Ha makes it a hard ceramic, refractory, with a melting point above 1600 C ^{[10] [11] [12]}.



FIGURE 1. Chemical structure of the Ha.

Ha's biomedical applications have been of great relevance due to their high degree of biocompatibility and bioactivity. Has been employed in the manufacture of different scaffold systems, including porous structures and hydrogels, with potential applications for the regeneration of bone tissue ^[13], and several methods have been developed to obtain this material, the main ones being precipitation, sol-gel, mechanochemistry, and solid-state reaction.

Ha can be obtained from natural sources, the most well-known are eggshells, corals, and animal bones, through thermal treatments at high temperatures ^[14]. Figure 2 provides a general description of the methodologies used to obtain Ha.



FIGURE 2. General scheme of the methodology to obtain Ha.

Biopolymers

The prefix "bio" indicates that these polymers are obtained from living matter or natural sources such as animals, plants, and microorganisms. They possess diverse characteristics such as biodegradability, low toxicity, biocompatibility, etc., which make them good candidates for a wide range of applications ranging from food and textiles up to electronics and medicine ^[15].

However, the term "biopolymer" is not strictly defined in the literature. There is some ambiguity in the term's biopolymers, biodegradable polymers, bio-based polymers and bioplastics ^[16]. Biopolymers are those polymers that are derived from natural resources and exhibit biodegradability, but do not include synthetic polymers that are biodegradable, they are also often referred to as natural polymers ^[17]. Biodegradable polymers include polymers of both natural and synthetic origin and are also known as bioresorbable polymers ^[18]. Bio-based polymers are those that are obtained partially or totally from renewable resources or waste and can be biodegradable or non-biodegradable ^[19]. And bioplastics can become biodegradable even if they are not necessarily derived from natural sources ^[20].

Biopolymers have been designed by nature itself with a single purpose, the sustenance of life. They can be grouped in different ways [21], as illustrated in Figure 3. Based on their nature, they are classified into three main categories: polysaccharides, proteins, and nucleic acids, where polysaccharides are macromolecular chains of low molecular weight that are linked by covalent sugar bonds, the best-known being glucose compounds or their derivatives such as starch and cellulose, which make the structure of plants. The proteins are the most important macromolecules present in animals, these are formed by the polymerization of amino acids, each type of protein has its own sequence of amino acids and structure which make its purpose determined, the most well-known proteins are hemoglobin, collagen, albumin, lactin, among others. Nucleic acids are linear polymers formed mainly of sugar molecules, phosphates, and bases, the most mentioned are RNA and DNA, which are found in living organisms for the purpose of storing and reading genetic information ^{[22] [23] [24].} Depending based on their polymeric structure are polysaccharides, polyesters, polypeptides, polyamides, polyurethanes, and vinyl polymers ^[25].



FIGURE 3. General information about biopolymers.

Due to the characteristics of biopolymers, they have become a tendency for their use in diverse applications, among which the pharmaceutical and biomedical areas stand out [22]. For this reason, the design of new materials based on biopolymers or biomaterials that include a composite of a polymer and a material in order to be compatible with human tissue, is one of the most promising areas in reconstructive medicine and orthopedics. In the case of implants made from biopolymers, they are used as a temporary osteoconductive scaffold, to which new bone tissue, compatible with the biopolymer, will eventually be anchored and grow. One of the greatest advantages of biopolymers in this area is that the implants do not require additional surgical intervention to remove them from the organism. Although one limitation they may have is their low mechanical properties since their high Young's modulus does not allow them to maintain the morphology of the polymeric implants under the load suffered by the musculoskeletal system ^[26] ^[27] ^[28]. Biopolymers, therefore, tend to have a wide field of utility, since they can be used for prostheses and implants, bone cements, and components of artificial organs ^[29].

Composites

Composites are materials consisting of two phases, illustrated in Figure 4, where the first is called the discontinuous or dispersed phase (reinforcement or filler) and the second is the continuous or "matrix" phase. The reinforcements or fillers can be fibers, particles, or sheets, whereas the matrix is the predominant phase ^[30].



FIGURE 4. Phases of a composite.

Composite materials have originated since old times, human beings have used them for different purposes, such as hunting weapons ^[31], in the same way nature provides these materials, for example, wood is composed of cellulose fibers in a lignin matrix, or the bones of mammals are constituted by collagen fibers in a calcium phosphate matrix ^[32].

These materials are developed to enhance their properties in relation to their pure components. The most desirable characteristics of these composites are their physicochemical properties, such as resistance to chemical attack, high mechanical properties, as well as favoring their use in everyday life. In the development of these composites, the aim has been to combine the properties of two or more components in a single compound, as well as to generate energy savings both in their production and application ^[33]. Based on this range of qualities, composites are considered as a feasible replacement for conventional materials in several industries such as aerospace, automotive, medical, among others ^[34].

Biopolymers and calcium phosphate salts composites have been used for bone replacement, because biopolymers are a good substitute for synthetic polymers, in addition to the fact that their composition can imitate the extracellular matrix due to the fact that they are bioabsorbable, biocompatible, and biodegradable. This type of materials with biomedical applications must have a great similarity with the structure and composition of the components present in the molecular structure of living beings; in this sense, proteins and polysaccharides are widely used in TE and regenerative medicine ^[35]. Therefore, the following sections will describe biopolymer/hydroxyapatite composites with potential biomedical applications, focusing on the most widely used for this purpose.

Alginate/Hydroxyapatite (Alg/Ha)

Alginic acid, also called alginate (Alg), occurs in nature mainly in brown algae (Phaeophyceae). Alg is an non-branched linear polysaccharide composed of the monomers β (1-4) D-mannuronic acid and α (1-4) L-guluronic acid, referred to as M- and G-block, respectively ^[36], its chemical structure is shown in Figure 5. Alg has diverse properties such as biodegradability, biocompatibility, bioadhesiveness, low toxicity, hydrophilicity, non-immunogenic effects, low cost, in addition to the fact that with the presence, even in low concentrations, of different cations (K⁺, Na⁺, Ca2⁺, Mg²⁺, Ba²⁺, and Sr²⁺) a stable hydrogel can be formed, due to the carboxylate groups of Alg ^[37]. In recent years, Alg has been reported to be utilized in medicine as a surgical and wound dressing ^[38], as a drug releasing agent and as a scaffold for bone, cartilage and/or skin ^[39]. These characteristics have led to it being widely studied as it is considered suitable for use in TE due to its biocompatibility with living cells and its property for the formation of hydrogels ^[40].



FIGURE 5. Chemical structure of sodium alginate.

With respect to Alg/Ha composites, these have been prepared by various processes and techniques, such as freeze-drying, electrospinning, extrusion, layer-bylayer, uniaxial hot pressing, liquid-gas systems, impregnation, coprecipitation ^[41], but the method most commonly used to obtaining them is crosslinking, however, it generally leads to a lack of homogeneity causing a decrease in mechanical properties ^[42].

Different researchers have developed this type of composites, such as that carried out in 2021 by Ocando *et al.*, who report obtaining an Alg/Ha composite in which Ha was doped with Mg, since it plays an important role in the proliferation of osteoblasts. In this work they obtained a biomaterial, by means of the cross-linking method, highly porous, with a uniform morphology, good biocompatibility and bioactivity ^[43].

This type of composite provides an improvement in different properties like mechanical strength, porosity, cell adhesion, biocompatibility, excellent osteogenic differentiation and mineralization ^[44], good bone regeneration, and can be used in the fixation of bone implants ^[45]. Table 1 shows the description of Alg/Ha systems where in some cases there are three-phase systems.

System	Possible applications	Properties/Results	Ref.
Alg/Ha-DS	Drug release	The <i>in-situ</i> production of Ha into the Alg polymeric matrix allowed for reduced swelling and dissolution rate. It was successfully applied to the release of the drug diclofenac sodium (DS) and a controlled release of up to 8 h was obtained.	[97]
Alg/Ha-CHX	Drug release in dentistry	The composite demonstrated good stability due to electrostatic interactions between chlorhexidine (CHX), Ha and Alg. In addition, a controlled release of CHX was achieved, obtaining the highest concentration at 72 h.	[98]
Alg/Ha	Implant	This composite developed was shown to have biological control within the subject under study, and the bone formed around the biomaterial had a lamellar structure, demonstrating that the biomaterial induces rapid bone production.	[99]
Alg/Ha	Biomedical application	The Alg/Ha composite tended to have better bioresorption at a higher weight percentage of Alg. An improvement in mechanical properties was also found compared to pure Ha and Alg.	[100]
Alg/Ha/MWCNT	Tissue engineering	The developed a composite with high porosity and showed improved compressive strength (72.02 ± 1.7 kPa). In addition, improved biocompatibility, differentiation and cell attachment were observed in the MG-63 cell line.	[101]
Alg/Ha	Tissue engineering	The obtained a composite that proved to have good biocompatibility and biodegradability. A histological study determined that the scaffolds developed can support both endochondral and intramembranous bone formation, and the defect was completely regenerated after 6 months.	[102]
Alg/Ha	Coatings	This composite showed improved mechanical properties and in microbiological studies they found that the composite significantly reduced the growth of the gram-positive bacterial <i>L. monocytogenes</i> compared to the pure Alg.	[103]

TABLE 1. Description and application of Alg/Ha systems.

Collagen/Hydroxyapatite (Col/Ha)

Collagen (Col), specifically type I, is the most abundant protein in the human body and can be found in different mammalian tissues such as the cornea, cartilage, skin, bones, tendons, ligaments, and others. It provides mechanical strength and stimulates cell adhesion and proliferation ^[36]. Is arranged as a triple helical structure which is mostly stabilized by hydrogen bridges ^[46] as shown in Figure 6. Has several properties that make it unique such as biodegradability, non-immunogenicity, bioabsorbability, and biocompatibility ^[47] ^[48].

Col has been used in TE and regenerative medicine for many years due to its biocompatibility and low immunogenicity, however, by itself it is not naturally osteoconductive and tends to have low compressive strength ^[49]. Col/Ha composites have been successfully



FIGURE 6. Chemical structure of collagen.

used in tissue procurement, since this combination provides good mechanical properties and osteoconductivity, in addition to favoring cell adhesion, cell migration, vascularization and osteoblastic differentiation ^[50]. In the same way, several researchers have opted for Col as the polymeric matrix par excellence in this type of composites since it is naturally present in the composition of human bones ^[51].

Col/Ha composites have been commonly obtained by different methods ^{[52] [53] [54]}:

- a) Mixing in collagen solution with Ha powder and subsequent lyophilization.
- b) Co-Precipitation of Ca and P precursors in collagen solution.
- c) Crosslinking for hydrogel formation.
- d) Mechanical mixing.
- e) Electrochemical deposition.

Studies realized in 2018 by Kaczmarek *et al.*, demonstrated that this type of composites shows an increment in osteoconductivity, mechanical properties, and biocompatibility ^[55], in the same sense these authors also found with this composite a high cell proliferation and a high mineralization potential ^{[56] [57]}.

Similarly, these biomaterials have a great potential to be applied in regenerative medicine through implants and/or bone tissue regenerator ^[58]. In addition, it can be used as a controlled drug release system thanks to its absorbent properties ^[59]. For a clearer comprehension, Table 2 presents a brief overview of Col/Ha systems.

System	Possible applications	Properties/Results	Ref.
Col/Ha	Tissue engineering	The composite has a bone composition similar to human bone and was shown to have superior mechanical properties compared to pure Col, thus providing reinforcement at the interphase.	[104]
Col/Ha	Tissue engineering	The developed composite showed to be doubled in strength with respect to its mechanical properties, thanks to the strengthening of the chemical bonds that occur between the Col-Ha, in addition, the scaffold maintained fibrillar structures and appropriate porosity, which permits a good cell proliferation.	[105]
Col/Ha/Graphene	Coatings	A composite with a composition very similar to that of human bone was developed and successfully electrodeposited on the surface of a Ti16Nb alloy. This coating provided a hydrophilic surface suitable for fibroblast cell development and adhesion.	[106]
Col/Ha	Bone defects	The composite produced adequately retains some trace elements that help its biocompatibility, thanks to the Ha synthesized from bovine bones, so it has a good bioactivity and can be used as a bone repair material.	[107]
Col/Ha/CNT	Bone replacement	This composite increased the flexural strength and fracture toughness up to 3%, and also exhibited positive osteoblast cell growth, thus improving its biological properties.	[108]
Col/Ha	Implant and bone regeneration	The composite exhibited a cell viability of 108.9 %, inferring that Ha and Col are biocompatible and non-toxic, since a viability above 100 % indicates cell proliferation.	[109]
Col/Ha	Tissue engineering	In the development of this composite obtained a high porosity of between 85-90 %, they also found an improvement in the compressive modulus obtaining a value over 1MPa, they also demonstrated that the architecture, permeability and composition of the scaffold favor infiltration and differentiation in vitro after subcutaneous implantation, proving that this type of scaffolds are osteoinductive.	[110]
Col/Ha	Tissue engineering	The composite shows good swelling properties, having that crosslinking and concentration of Col and Ha is important to modify the microstructure, mechanical properties and degradation rate of the composite.	[111]

TABLE 2. Description of Col/Ha systems.

Gellan Gum/Hydroxyapatite (GG/Ha)

Natural gums can be obtained from different sources such as vegetable fibers (gum arabic, gum ghatti, gum tragacanth, etc.), seeds (guar gum, konjac gum, etc.), algae (agar-agar gum, alginates, etc.), or microorganisms (gellan gum, xanthan gum, rhamsan gum, etc.) ^[60].

Microbial gums are of high molecular weight, approximately 1,000,000 Da, and are composed of carbohydrates, proteins, lipids, and humic substances. Are produced by certain microorganisms from the aerobic fermentation of sugar ^[61], Gellan gum (GG) is an exopolysaccharide of anionic nature consisting of a repeating unit of a tetrasaccharide (D-glucose, 1-3, glucuronic acid, D-glucose and 1,4, L-rhamnose), see Figure 7.



FIGURE 7. Chemical structure of gellan gum.

Similar to Alg, GG forms gels in presence of metal ions and can be easily processed ^[36], some of the main characteristics of GG are its non-toxicity, biocompatibility, and biodegradability.

This polysaccharide is produced by the excretion of the bacteria *Sphingonomas elodea* and *Sphingomonas pauci-mobilis*. In addition, is highly thermally stable and the gels obtained are mainly translucent ^[62] and have high porosity, antibacterial nature, proliferation, and bone formation ^[63]. They are commonly prepared by lyophilization, rehydration, and co-precipitation for the formation of cellular adhesive sponge-type hydrogels ^[64]. Diverse studies have shown that the gelation temperature, strength, texture, clarity, and speed of gel formation depend directly on the pH value during the obtaining process ^[60].

Due to its characteristics, GG has been used in diverse applications in the food, cosmetic and pharmaceutical industries; in the last of these, its application in ophthalmology, drug release, cell immobilization, biosensor synthesis, among others, stands out ^[64]. In recent decades, GG has been studied for numerous applications in cellular tissue, including brain TE, dentistry, cartilage, and intervertebral disc ^[65]. Shin *et al.* employed the photoreticulation reaction in order to obtain rigid microgels of GG. The product obtained was immersed in a modified gelatin solution, which was photoreticulated, and the final material showed increased mechanical strength ^[66].

On the other hand, several studies report that it is possible to increase the mechanical strength of GG by adding calcium phosphate salts, such as Ha. These inorganic particles can be easily incorporated into hydrogels by mechanical mixing ^[67]. Previous studies have shown that mineralization of GG hydrogels could enhance mechanical properties, as well as promote cell adhesion and growth, and osteogenic differentiation ^[68].

Recent research by Pereira *et al.* at 2018 successfully reported the obtaining of a GG/Ha composite by incorporating different concentrations of Ha (5, 10, 15, and 20 %) using the crosslinking method, obtaining a material with improved mechanical properties, high stability in degradation and promoting high cell proliferation, null toxicity, and excellent biocompatibility ^[69]. The Table 3 shows the description of GG/Ha composites.

Chitosan/Hydroxyapatite (Ch/Ha)

Chitin is the second most abundant organic compound on planet earth, it is a biopolymer found in various living organisms, such as the animal, plant, and fungi kingdoms. It is found in the exoskeleton of crustaceans, the cuticle of insects, and cell walls of fungi, for example, in shrimp, lobsters, crabs, squid, mollusks, scorpions, ants, beetles, cockroaches, bees, spiders, *Agaricus bisporus, Aspergillus niger, Auricularia auriculajudae*, and others. Chitin is an effective material for suturing, wound heal-

System	Possible applications	Properties/Results	Ref.
GG/Ha	Bone repairment	The addition of Ha in the polymeric matrix of GG is reported to favor mechanical strength.	[112]
GG/Ha	Bone regeneration	This composite was made in 3D which allowed a maturation of the bone cells because it assimilates the structure of human bone.	[113]
GG/Ha	Bone reconstruction	The obtained composites showed high mechanical strength, good porosity with the capacity to absorb nutrients necessary for proper cell growth, as well as slow degradation, which provides the necessary time to induce osteoconduction and the generation of new bone.	[114]
GG/Ha	Bone replacements	The addition of Ha in the GG polymer matrix provides good cell adhesion on the surface of the material. in addition, it was observed that increasing the percentage of Ha significantly increased the proliferation of MC3T3- E1 cells.	[115]
GG/Ha	Bone grafting	The developed material improved its mechanical properties with respect to pure GG, in addition to showing a clear organization of the cytoskeleton and cell motility from day 7. Hence, this composite favors cell adhesion and propagation.	[116]
GG/Ha-Ce	Bone grafting	A composite of GG/Ha developed with the addition of doped cerium (Ce) to the hydroxyapatite. This material demonstrated high porosity, increased mechanical properties, and demonstrated that it is non-toxic and biocompatible.	[117]

TABLE 3. Description of GG/Ha systems	TABLE 3.	Descri	ption of	GG/	Ha sy	stems.
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ing and drug release thanks to its biocompatibility, non-toxicity, immunogenicity, biodegradability and antimicrobial activity [70]. Although the most important aspect of chitin is its main derivative, chitosan (Ch), which is obtained by deacetylation of chitin. Chemically, Ch has a similar structure to cellulose and chitin, as shown in Figure 8, its differentiator is the functional group at the C-2 position. While cellulose has a hydroxyl group (-OH) and chitin has an N-acetylamine group (-NHCOOH3), Ch has an amino group (-NH2). Ch is a linear polysaccharide consisting of an acetylated unit (N-acetyl-2-amino-2deoxy-d-glucopyranose) and a deacetylated unit (2-amino-2-deoxy-glucopyranose), where the repetitive units are linked by $\beta(1-4)$ glycosidic bonds ^[71]. The degree of deacetylation and molecular weight are of major influence on the physicochemical properties of Ch, where the degree of deacetylation represents the ratio of glucosamine to N-acetyl-glusoamine [72].

Non-toxicity, biodegradability, biocompatibility, antimicrobial activity, anticancer activity, antiinflammatory, hydrophilicity, crystallinity, immunogenicity, cell



adhesion, cell proliferation and differentiation, and osteoconductivity ^{[73] [74] [75]} are some of the properties of Ch which make it unique. For this reason, it has been used in countless applications in the food industry, agriculture, pharmacy, medicine, dentistry, ophthalmology, cosmetics, the textile industry, veterinary medicine, wastewater treatment, and has also been used in the formulation of fire-retardant foams, among others ^{[71] [76] [77]}. Ch has been shown to be suitable for use in IT as a bone tissue graft, in the manufacture of porous scaffolds, films for superficial wounds, drug release, and in formulations for the production of hydrogels ^[78].

However, Ch presents as a disadvantage its poor mechanical properties [79], so to improve this, several investigations report the development of composites based on Ch/Ha because both biomaterials present excellent biocompatibility with human cellular tissue ^[80], it is reported that this composite presents improvements in its mechanical properties, due to the binding of the amino and hydroxyl groups of Ch to the calcium ions present in Ha^{[81][82]}. The Ch/Ha composite can be produced mainly by freeze-drying; this technique is one of the most widely used to obtain scaffolds, and other techniques can also be used, such as NSN textile methodology for the production of fibrous structures and the use of enzymes. This composite can be used as a bioactive interface between the metallic implant and the human bone [75]. In 2017 Okada et al., used the coagulation method to obtain a Ch/Ha composite fiber, where they reported that this material improves the mechanical properties [83]. Also, Balagangadharan et al., reported in 2018 the development of a Ch/Ha composite to which they added ZrO2 nanoparticles obtaining scaffolds, which presented an osteoconductive improvement and osteoblast differentiation, resulting in a beneficial composite for bone tissue regeneration [84]. Descriptions of some of the Ch/Ha systems are presented in Table 4.

Polylactic Acid-Hydroxyapatite (PLLA/Ha)

Polylactic acid (PLLA), its chemical structure can be seen in Figure 9, is a linear aliphatic thermoplastic polyester derived 100% from renewable resources with wide availability of raw materials (sugar, corn, potatoes, sugarcane, sugar beet, among many others) ^[85]. This material is simple to process, using any technique, such as extrusion, injection and compression molding, thermoforming and foaming, as well as blow molding, electrospinning, spinning, and stretching of molten polymer, among others. Extrusion is commonly used for PLA processing because it can be adapted for various applications ^[86], The industry most known is packaging; in fact, it is approved by the United States Food and Drug Administration (FDA) ^[87], for its excellent properties of transparency, low toxicity, versatility, osteoconductivity, biodegradability, biocompatibility, bioresorbability ^[88], properties influenced by its molecular weight and stereochemistry ^[89].



FIGURE 9. Chemical structure of polylactic acid.

PLLA exists in two stereoisomeric forms D-PLLA (DPLLA) and L-PLLA (PLLA), and a mixture of D- and L-lactic acid also exists as D,L-PLLA (PDLLA). Polymers derived from the optically active D- and L-monomers are semi-crystalline and those derived from the optically inactive D,L-PLLA are amorphous. Generally, L-PLLA is preferred for applications where high mechanical strength and toughness are required, for example, orthopedic devices and sutures. On the other hand, due to the amorphous nature of D,L-PLLA is considered for drug release applications, since for this purpose it is very important to have a homogeneous dispersion of the active species within a matrix. It has been reported that PLLA degradation is significantly affected with increasing crystallinity, so the mixture of PLLA and PDLLA is effective in adjusting the crystallization and morphology of the polymer, and thus its mechanical properties [90]. In recent years, it has been successfully used as a surgical implant material, drug release, and in the fabrication of 3D printed scaffolds [91].

System	Possible applications	Properties/Results	Ref.
Ch/Ha/Magnetite	Superficial wounds	The Ch/Ha composite increases the bending modulus up to 2.6 GPa, with the addition of magnetite, however, it showed a lower bending strength. In contrast, magnetic nanoparticles had no negative effect on the cytocompatibility of the material.	[118]
Ch/Ha	Coatings	The composites developed showed good swelling and crosslinking properties, and the L929 and Saos-2 cell lines showed cell proliferation.	[119]
Ch/Ha-DOX	Drug release	This study incorporated an anticancer drug (doxorubicin, DOX) into Ha nanoparticles modifying this with Ch. The modified Ha did not show cytotoxicity in the MG-63 cell line.	[120]
Cu-Ch/Ha-Sr	Tissue engineering	The composite showed an improvement in the mechanical properties of Ch, as well as excellent cell infiltration, cell migration and neovascularization due to the high porosity, making them cytocompatible.	[121]
MWCNT-Ch/Ha	Bone replacements	The nanocomposite exhibited excellent mechanical properties, good biocompatibility and bioactivity, hence fulfilling the required properties of the extracellular matrix of bone.	[122]
Ch/Ha	Bone replacements and coatings	The composite had a tensile strength of 30-70 MPa, showed to be cytocompatible in the L929 cell line after 24 h of incubation.	[123]
Ch/HaNPsAg	Drug release and implant	This type of scaffolds exhibited an improvement in mechanical resistance, as well as improved cell penetration, adhesion and propagation. The presence of silver nanoparticles (NPsAg) provides antibacterial activity, thus reducing the risk of infections.	[124]
Ch/Ha	Tissue engineering	The composite showed a high similarity compared to natural bone. Also observed that increasing the concentration of Ha resulted in a higher adhesion of osteoblasts. In addition, this material showed a high degree of proliferation in the MC3T3-E1 cell line. Compared to the pure Ch scaffold, the compressive strength increased by 33.07 %.	[125]

A disadvantage that PLLA may present is the absence of the property that facilitates cell binding on its surface and its proliferation, due to a scarcity of cell binding capacity ^[92]. This deficiency in cohesion could be improved by employing the addition of Ha as a strategy since calcium phosphates promote cell proliferation on the scaffold surface. It has also been shown that PLA/ Ha composites tend to enhance their overall properties such as biocompatibility, resorbability, osteoconductivity, cell proliferation, and mechanical strength ^[93].

The dispersion of Ha on PLA is considered one of the critical factors that determine the properties of the composite, for which different methods and techniques are used to obtain it, highlighting melt mixing, electrospinning, processing by compression, emulsion and solvent casting, the latter being the most used ^[94]. On the other hand, Zhang *et al.*, reported in 2015 the obtaining of a

PLA/Ha composite by the electrospinning method, which led to an easily obtained material with great potential in terms of its biomedical applications such as rapid mineralization and high biocompatibility ^[95]. Also, at 2019 de Moura *et al.*, obtained a PLA/Ha material with improved mechanical resistance and bioactivity, reported that the addition of Ha increases the protein adsorption capacity, suppresses cell death by apoptosis, and creates a favorable microenvironment for cell proliferation ^[96]. The table 5 present a description of the PLLA/Ha systems and their potential application.

CONCLUSIONS

Biopolymers are materials used in a diversity of applications due to their excellent qualities such as biodegradability, biocompatibility, and non-toxicity, therefore several researchers have studied them for their application in tissue engineering, however, these materials have poor

System	Possible applications	Properties/Results	Ref.
PLLA/Ha	Bone replacements	The composite had an increase in Young's modulus incorporating a low quantity of Ha in the polymeric matrix. It was also demonstrated that the material is not cytotoxic in the MC3T3-E1 cell line, in besides having a good osteoblastic maturation.	[126]
PLLA/Ha	Bone tissue engineering	The developed composite has a hydrophilic part of the Ha, which contributes to cell growth and bone repair, as well as a hydrophobic part of the PLLA, which acts as a barrier between tissues. As a result, this material has a high cytocompatibility.	[127]
PLLA/Ha- NPsAg	Coatings	The antimicrobial activity of the composite exhibited better inhibition efficacy on <i>E. coli</i> bacteria. In the hemocompatibility assay showed 2 % hemolytic activity, so it is considered compatible with human blood.	[128]
PLLA/Ha	Coatings	The composite shown that the mechanical properties and thermal behavior increased with the addition of Ha to the PLLA matrix. In addition, the material exhibited good biocompatibility.	[129]
PLLA/Ha	Scaffold	The composite demonstrated mechanical properties comparable to those of normal bone. Furthermore, it exhibited excellent 3D printability at 90:10 ratio of PLLA/Ha respectively.	[130]
PLLA/Ha	Scaffold	The composite demonstrated good adhesion and cellular proliferation in the material, with adequate osteoconductivity.	[131]
PLLA/Ha	Scaffold	The composite showed good adhesion and cell proliferation on the surface, adequate porosity, as well as an increase in mechanical properties with respect to pure PLLA.	[132]

TABLE 5. Description of PLLA/Ha systems.

mechanical properties. In response to this mechanical deficiency, composites based on biopolymers/calcium phosphate salts (hydroxyapatite) have been used, since it has been shown that these types of materials show improvements in their mechanical properties and biocompatibility. For this reason, these composites are a viable option for the development of biomaterials that can be used in different applications in tissue engineering, such as drug delivery, tissue implants, scaffolds, etc.

AUTHOR CONTRIBUTIONS

J.D.F.V. conceptualized the project, performed bibliographical research, and provided briefing to the team members to carry out the project, participated in the writing of the original manuscript and translated it to the English language. A.S.G. provided funding through a SEP-CONACYT grant for basic science, specially used for providing payment for publication; oversaw the scheduling and progress of the project; contributed to the writing of the manuscript, mainly with comments from her field of expertise. C.M.L.B. contributed to the writing of the manuscript, especially with topics regarding her field of expertise, provided bibliographical resources key to the finalization of the project and helped with the English translation. A.O.C.F. contributed to writing the draft and final version of the manuscript, edited comments on her field of expertise key to the conclusion of the project. P.A.V. conceptualized the project, designed the objectives of the project, and oversaw its development; participated in all the writing stages of the manuscript and helped with the translation to English language. All authors reviewed and approved the final version of the manuscript.

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CONFLICTS OF INTEREST

The authors do not report any conflict of interest.

REFERENCES

- El-Habashy SE, Eltaher HM, Gaballah A, Zaki EI, et al. Hybrid bioactive hydroxyapatite/polycaprolactone nanoparticles for enhanced osteogenesis. Mater Sci Eng C [Internet].
 2021;119:111599. Available from: https://doi.org/10.1016/j.msec.2020.111599
- [2] Escobar Sierra D, Mesa Ospina D. Evaluación de recubrimientos de quitosano sobre cuerpos porosos de hidroxiapatita. Sci Tech [Internet]. 2019;24(1):161-172. Available from: https://doi.org/10.22517/23447214.20051
- [3] Chen J, Yu Q, Zhang G, Yang S, et al. Preparation and biocompatibility of nanohybrid scaffolds by in situ homogeneous formation of nano hydroxyapatite from biopolymer polyelectrolyte complex for bone repair applications. Colloids Surf B [Internet]. 2012;93:100-107. Available from: http://dx.doi.org/10.1016/j.colsurfb.2011.12.022
- [4] Chakravarty J, Rabbi MF, Chalivendra V, Ferreira T, et al. Mechanical and biological properties of chitin/polylactide (PLA)/ hydroxyapatite (HAP) composites cast using ionic liquid solutions. Int J Biol Macromol [Internet]. 2020;151:1213-1223. Available from: https://doi.org/10.1016/j.ijbiomac.2019.10.168
- [5] Kalisz G, Przekora A, Kazimierczak P, Gieroba B, et al. Physicochemical changes of the chitosan/β-1,3-glucan/ hydroxyapatite biocomposite caused by mesenchymal stem cells cultured on its surface in vitro. Spectrochim Acta A Mol Biomol [Internet]. 2021;251:119439. Available from: https://doi.org/10.1016/j.saa.2021.119439
- [6] Benedini L, Laiuppa J, Santillán G, Baldini M, et al. Antibacterial alginate/nano-hydroxyapatite composites for bone tissue engineering: Assessment of their bioactivity, biocompatibility, and antibacterial activity. Mater Sci Eng C [Internet]. 2020;115:111101. Available from: https://doi.org/10.1016/j.msec.2020.111101
- Sathiyavimal S, Vasantharaj S, LewisOscar F, Selvaraj R, Brindhadevi K, Pugazhendhi A. Natural organic and inorganichydroxyapatite biopolymer composite for biomedical applications. Prog Org Coat [Internet]. 2020;147:105858. Available from: https://doi.org/10.1016/j.porgcoat.2020.105858
- [8] Lett AJ, Sagadevan S, Fatimah I, Hoque ME, et al. Recent advances in natural polymer-based hydroxyapatite scaffolds: Properties and applications. Eur Polym [Internet]. 2021;148:110360. Available from: https://doi.org/10.1016/j.eurpolymj.2021.110360
- [9] Trakoolwannachai V, Kheolamai P, Ummartyotin S. Characterization of hydroxyapatite from eggshell waste and polycaprolactone (PCL) composite for scaffold material. Compos B Eng [Internet]. 2019;173:106974. Available from: https://doi.org/10.1016/j.compositesb.2019.106974
- [10] Williams RAD, Elliot JC. Bioquímica dental básica y aplicada. Distrito Federal, México: El Manual Moderno; 1990. 263p. Spanish.
- [11] Rahavi SS, Ghaderi O, Monshi A, Fathi MH. A comparative study on physicochemical properties of hydroxyapatite powders derived from natural and synthetic sources. Russ J Non-Ferrous Metals [Internet]. 2017;58(3):276-286. Available from: https://doi.org/10.3103/S1067821217030178
- [12] Giraldo-Betancur A, Espinosa-Arbelaez DG, del Real-López A, Millan-Malo BM, et al. Comparison of physicochemical properties of bio and commercial hydroxyapatite. Curr Appl Phys [Internet]. 2013;13(7):1383-1390. Available from: http://dx.doi.org/10.1016/j.cap.2013.04.019

- [13] Alvarez-Barreto J, Márquez K, Gallardo E, Moret J, et al. Mesenchymal Stem Cell Culture on Composite Hydrogels of Hydroxyapatite Nanoparticles and Photo-Crosslinking Chitosan. Rev Mex Ing Biom [Internet]. 2017;38(3):524-536. Available from: http://dx.doi.org/10.17488/RMIB.38.3.2
- [14] Rivera JA, Fetter G, Bosch P. Efecto del pH en la síntesis de hidroxiapatita en presencia de microondas. Rev Mater [Internet]. 2011;15(4):488-505. Available from: https://doi.org/10.1590/51517-70762010000400003
- [15] Flores-Valdez JD, Sáenz-Galindo A, Múzquiz-Ramos EM, Soria Aguilar MJ. Biopolymers and applications. CienciAcierta [Internet]. 2021;(66):61-72. Available from: http://www.cienciacierta.uadec. mx/articulos/CC66/biopolimerosyaplicaciones.pdf
- [16] Sivakanthan S, Rajendran S, Gamage A, Madhujith T, et al. Antioxidant and antimicrobial applications of biopolymers: A review. Food Res Int [Internet]. 2020;136:109327. Available from: https://doi.org/10.1016/j.foodres.2020.109327
- [17] Lizundia E, Kundu D. Advances in Natural Biopolymer-Based Electrolytes and Separators for Battery Applications. Adv Funct [Internet]. 2021;31(3):2005646. Available from: https://doi.org/10.1002/adfm.202005646
- [18] Hochmańska-Kaniewska P, Janiszewska D, Oleszek T. Enhancement of the properties of acrylic wood coatings with the use of biopolymers. Prog Org Coat [Internet]. 2022;162:106522. Available from: https://doi.org/10.1016/j.porgcoat.2021.106522
- [19] Tuan Naiwi TSR, Aung MM, Rayung M, Ahmad A, et al. Dielectric and ionic transport properties of bio-based polyurethane acrylate solid polymer electrolyte for application in electrochemical devices. Polym Test [Internet]. 2022;106:107459. Available from: https://doi.org/10.1016/j.polymertesting.2021.107459
- [20] Wan Mahari AW, Kee SH, Foong SY, Amelia TSM, et al. Generating alternative fuel and bioplastics from medical plastic waste and waste frying oil using microwave co-pyrolysis combined with microbial fermentation. Renew Sustain Energy Rev [Internet]. 2022;153:111790. Available from: https://doi.org/10.1016/j.rser.2021.111790
- [21] Parveen FK. Recent Advances in Biopolymers [Internet]. London: IntechOpen; 2016. 288p. Available from: https://doi.org/10.5772/60630
- [22] Narain, R. Polymer Science and Nanotechnology Fundamentals and Applications [Internet]. United Kingdom: Elsevier, 2020. 488p. Available from: <u>https://doi.org/10.1016/C2018-0-01134-2</u>
- [23] Rebelo R, Fernandes M, Fangueiro R. Biopolymers in Medical Implants: A Brief Review. Procedia Eng [Internet]. 2017;200:236-243. Available from: https://doi.org/10.1016/j.proeng.2017.07.034
- Hassan MES, Bai J, Dou D-Q. Biopolymers; Definition, Classification and Applications. Egypt J Chem [Internet]. 2019;62(9):133-145.
 Available from: https://dx.doi.org/10.21608/ejchem.2019.6967.1580
- [25] Ahmed S, Kanchi S, Kumar G. Handbook of Biopolymers: Advances and Multifaceted Applications. California: Jenny Stanford Publishing; 2018. 308p.
- [26] Dubinenko GE, Zinoviev AL, Bolbasov EN, Novikov VT, et al. Preparation of Poly(L-lactic acid)/Hydroxyapatite composite scaffolds by fused deposit modeling 3D printing. Mater Today Proc [Internet]. 2020;22:228-234. Available from: https://doi.org/10.1016/j.matpr.2019.08.092

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- [27] Kumar PPP, Lim D-K. Gold-Polymer Nanocomposites for Future Therapeutic and Tissue Engineering Applications. Pharmaceutics [Internet]. 2021;14(1):70. Available from: https://doi.org/10.3390/pharmaceutics14010070
- [28] Biswal T. Biopolymers for tissue engineering applications: A review. Mater Today Proc [Internet]. 2021;41:397-402. Available from: <u>https://doi.org/10.1016/j.matpr.2020.09.628</u>
- [29] Villareal Valdiviezo GP, Múzquiz Ramos EM, Farías Cepeda L. Medical applications of biopolymers. CienciAcierta [Internet]. 2020;63:83. Available from: http://www.cienciacierta.uadec.mx/ articulos/CC63/83AplicacionesMedicas.pdf
- [30] Aboudi J, Arnold SM, Bednarcyk BA. Mechanics of Composite Materials [Internet]. 2nd ed. Butterworth-Heinemann: Elsevier; 2013. 984p. Available from: https://doi.org/10.1016/C2011-0-05224-9
- [31] Gay D, Hoa SV, Tsai SW. Composite Materials: Design and Applications. 1st ed. Florida: CRC Press; 2002. 552p.
- [32] Gibson RE. Principal of Composite Mechanics. 4th ed. Boca Raton: CRC Press; 2016. 700p.
- [33] Mohanty AK, Misra M, Drzal LT, Selke S, Harte B, Hinrichsen G. Natural Fibers, Biopolymers, and Biocomposites [Internet]. 1st ed. Boca Raton: CRC Press; 2005. 896p. Available from: https://doi.org/10.1201/9780203508206
- [34] Sankaran S, Ravishankar BN, Ravi Sekhar K, Dasgupta S, et al. Syntactic Foams for Multifunctional Applications. In: Kar, K (eds.). Composite Materials [Internet]. Berlin: Springer Berlin Heidelberg; 2017. 281-314p. Available from: https://doi.org/10.1007/978-3-662-49514-8 9
- [35] Holban AM, Grumezescu A (eds.). Materials for Biomedical Engineering: Hydrogels and Polymer-based Scaffolds [Internet]. Amsterdam, Oxford, Cambridge: Elsevier; 2019. 562p. Available from: https://doi.org/10.1016/C2017-0-04477-4
- [36] Moraes MA, Silva CF, Vieira RS (eds.). Biopolymers Membranes and Films Health, Food, Environment, and Energy Applications. Amsterdam, Oxford, Cambridge: Elsevier; 2020. 633p. Available from: https://doi.org/10.1016/C2018-0-02693-6
- [37] Benedini L, Laiuppa J, Santillán G, Baldini M, et al. Antibacterial alginate/nano-hydroxyapatite composites for bone tissue engineering: Assessment of their bioactivity, biocompatibility, and antibacterial activity. Mater Sci Eng C [Internet]. 2020;115:11101. Available from: https://doi.org/10.1016/j.msec.2020.111101
- [38] Wang L, Li Y, Li C. In situ processing and properties of nanostructured hydroxyapatite/alginate composite. J Nanopart Res [Internet]. 2009;11(3):691-699. Available from: https://doi.org/10.1007/s11051-008-9431-y
- [39] Chae T, Yang H, Leung V, Ko F, et al. Novel biomimetic hydroxyapatite/alginate nanocomposite fibrous scaffolds for bone tissue regeneration. J Mater Sci: Mater Med [Internet]. 2013;24(8):1885-1894. Available from: https://doi.org/10.1007/s10856-013-4957-7
- [40] Sukhodub LF, Sukhodub LB, Litsis O, Prylutskyy Y. Synthesis and characterization of hydroxyapatite-alginate nanostructured composites for the controlled drug release. Mater Chem Phys [Internet]. 2018;217:228-34. Available from: https://doi.org/10.1016/j.matchemphys.2018.06.071
- [41] Mahmoud EM, Sayed M, El-Kady AM, Elsayed H, et al. In vitro and in vivo study of naturally derived alginate/hydroxyapatite bio composite scaffolds. Int J Biol Macromol [Internet]. 2020;165:1346-1360. Available from: https://doi.org/10.1016/j.ijbiomac.2020.10.014

- [42] You F, Chen X, Cooper DML, Chang T, et al. Homogeneous hydroxyapatite/alginate composite hydrogel promotes calcified cartilage matrix deposition with potential for three-dimensional bioprinting. Biofabricación [Internet]. 2018;11:015015. Available from: https://doi.org/10.1088/1758-5090/aaf44a
- [43] Ocando C, Dinescu S, Samoila I, Ghitulica CD, et al. Fabrication and properties of alginate-hydroxyapatite biocomposites as efficient biomaterials for bone regeneration. Eur Polym [Internet]. 2021;151:110444. Available from: https://doi.org/10.1016/j.eurpolymj.2021.110444
- [44] Sukhodub LF, Sukhodub LB, Pogrebnjak AD, Turlybekuly A, et al. Effect of magnetic particles adding into nanostructured hydroxyapatite-alginate composites for orthopedics. J Korean Ceram Soc [Internet]. 2020;57(5):557-569. Available from: https://doi.org/10.1007/s43207-020-00061-w
- [45] De Paula FL, Barreto IC, Rocha-Leão MH, Borojevic R, et al. Hydroxyapatite-alginate biocomposite promotes bone mineralization in different length scales in vivo. Front Mater Sci China. 2009;3(2):145-153. Available from: https://doi.org/10.1007/s11706-009-0029-9
- [46] González Paz R, Grillo A, Feijoo JL, Noris-Suárez K, et al. Estudio de Mezclas de Polietileno de Alta Densidad (PEAD) con colágeno/acetato de sodio e Hidroxiapatita (HA). In: Müller-Karger C, Wong S, La Cruz A. (eds). IV Latin American Congress on Biomedical Engineering 2007 [Internet]. Berlin: IFMBE Proceedings; 2008;18:676-680. Available from: https://doi.org/10.1007/978-3-540-74471-9_157
- Yoruc ABH, Aydınoglu AK. Synthesis of Hydroxyapatite/Collagen (HA/COL) Composite Powder Using a Novel Precipitation Technique. Acta Phys Pol [Internet]. 2015;127(4):1264-1267. Available from: http://dx.doi.org/10.12693/APhysPolA.127.1264
- [48] Sukul M, Min Y-L, Lee B-T. Collagen-hydroxyapatite coated unprocessed cuttlefish bone as a bone substitute. Mater [Internet]. 2016;181:156-560. Available from: http://dx.doi.org/10.1016/j.matlet.2016.05.170
- [49] Lara-Rico R, Claudio-Rizo JA, Múzquiz-Ramos E, Lopez-Badillo CM. Hidrogeles de colágeno acoplados con hidroxiapatita para aplicaciones en ingeniería tisular. TIP Rev Espec Cienc Quim-Biol [Internet]. 2020;23:1-12. Available from: https://doi.org/10.22201/fesz.23958723e.2020.224
- [50] Cholas R, Padmanabhan SK, Gervaso F, Udayan G, et al. Scaffolds for bone regeneration made of hydroxyapatite microspheres in a collagen matrix. Mater Sci Eng C [Internet]. 2016;63:499-505. Available from: http://dx.doi.org/10.1016/j.msec.2016.03.022
- [51] Bhuiyan D, Jablonsky MJ, Kolesov I, Middleton J, et al. Novel synthesis and characterization of a collagen-based biopolymer initiated by hydroxyapatite nanoparticles. Acta Biomater [Internet]. 2015;15:181-190. Available from: http://dx.doi.org/10.1016/j.actbio.2014.11.044
- [52] Sun R-X, Lv Y, Niu Y-R, Zhao X-H, et al. Physicochemical and biological properties of bovine-derived porous hydroxyapatite/ collagen composite and its hydroxyapatite powders. Ceram Int [Internet]. 2017;43(18):16792-16798. Available from: http://dx.doi.org/10.1016/j.ceramint.2017.09.075
- [53] Becerra J, Rodriguez M, Leal D, Noris-Suarez K, et al. Chitosancollagen-hydroxyapatite membranes for tissue engineering. J Mater Sci: Mater Med [Internet]. 2022;33(2):18. Available from: https://doi.org/10.1007/s10856-022-06643-w

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- [54] Zhao X, Li H, Xu Z, Li K, et al. Selective preparation and characterization of nano-hydroxyapatite/collagen coatings with three-dimensional network structure. Surf Coat Technol [Internet]. 2017;322:227-237. Available from: http://dx.doi.org/10.1016/j.surfcoat.2017.05.042
- [55] Kaczmarek B, Sionkowska A, Osyczka AM. Physicochemical properties of scaffolds based on mixtures of chitosan, collagen and glycosaminoglycans with nano-hydroxyapatite addition. Int J Biol Macromol [Internet]. 2018;118:1880-1883. Available from: https://doi.org/10.1016/j.ijbiomac.2018.07.035
- [56] Kaczmarek B, Sionkowska A, Gołyńska M, Połkowska I, et al. In vivo study on scaffolds based on chitosan, collagen, and hyaluronic acid with hydroxyapatite. Int J Biol Macromol [Internet]. 2018;118:938-944. Available from: https://doi.org/10.1016/j.ijbiomac.2018.06.175
- [57] Kikuchi M, Itoh S, Ichinose S, Shinomiya K, et al. Self-organization mechanism in a bone-like hydroxyapatite/collagen nanocomposite synthesized in vitro and its biological reaction in vivo. Biomaterials [Internet]. 2001;22(13):1705-1711. Available from: https://doi.org/10.1016/S0142-9612(00)00305-7
- [58] Dou DD, Zhou G, Liu HW, Zhang J, et al. Sequential releasing of VEGF and BMP-2 in hydroxyapatite collagen scaffolds for bone tissue engineering: Design and characterization. Int J Biol Macromol [Internet]. 2019;123:622-628. Available from: https://doi.org/10.1016/j.ijbiomac.2018.11.099
- [59] Taniyama T, Masaoka T, Yamada T, Wei X, et al. Repair of Osteochondral Defects in a Rabbit Model Using a Porous Hydroxyapatite Collagen Composite Impregnated With Bone Morphogenetic Protein-2. Artif Organs [Internet]. 2015;39(6):529-535. Available from: https://doi.org/10.1111/aor.12409
- [60] Osmałek T, Froelich A, Tasarek S. Application of gellan gum in pharmacy and medicine. Int J Pharm. 2014;466(1-2):328-340. Available from: https://doi.org/10.1016/j.ijpharm.2014.03.038
- [61] Zare EN, Makvandi P, Borzacchiello A, Tay FR, et al. Antimicrobial gum bio-based nanocomposites and their industrial and biomedical applications. Chem Commun [Internet]. 2019;55(99):14871-14885. Available from: https://doi.org/10.1039/C9CC082076
- [62] Santos MVB, Oliveira AL, Osajima JA, Silva-Filho EC. Development of composites scaffolds with calcium and cerium-hydroxyapatite and gellan gum. Ceram Int [Internet]. 2020;46(3):3811-3817. Available from: https://doi.org/10.1016/j.ceramint.2019.10.104
- [63] Rajesh R, Ravichandran YD, Reddy MJK, Ryu SH, et al. Development of functionalized multi-walled carbon nanotubebased polysaccharide-hydroxyapatite scaffolds for bone tissue engineering. RSC Adv [Internet]. 2016;6(85):82385-82393. Available from: http://dx.doi.org/10.1039/C6RA16709H
- [64] Manda MG, da Silva LP, Cerqueira MT, Pereira DR, et al. Gellan gum-hydroxyapatite composite spongy-like hydrogels for bone tissue engineering. J Biomed Mater Res A [Internet]. 2018;106(2):479-490. Available from: https://doi.org/10.1002/jbm.a.36248
- [65] Vieira S, da Silva Morais A, Garet E, Silva-Correia J, et al. Selfmineralizing Ca-enriched methacrylated gellan gum beads for bone tissue engineering. Acta Biomater [Internet]. 2019;93:74-85. Available from: https://doi.org/10.1016/j.actbio.2019.01.053
- [66] Shin H, Olsen BD, Khademhosseini A. Gellan gum microgelreinforced cell-laden gelatin hydrogels. J Mater Chem B [Internet]. 2014;2(17):2508-2516. Available from: https://doi.org/10.1039/C3TB20984A

- [67] Nayak AK, Alkahtani S, Hasnain MS. Jackfruit Seed Starch-Based Composite Beads for Controlled Drug Release. In: Nayak AK, Alkahtani S, Hasnain MS (eds). Polymeric and Natural Composites. Advances in Material Research and Technology [Internet]. Springer Cham ;2022. 213-240p. Available from: https://doi.org/10.1007/978-3-030-70266-3_7
- [68] Xu L, Bai X, Yang J, Li J, et al. Preparation and characterisation of a gellan gum-based hydrogel enabling osteogenesis and inhibiting Enterococcus faecalis. Int J Biol Macromol [Internet].
 2020;165:2964-2973. Available from: https://doi.org/10.1016/j.ijbiomac.2020.10.083
- [69] Pereira DR, Canadas RF, Silva-Correia J, da Silva Morais A, et al. Injectable gellan-gum/hydroxyapatite-based bilayered hydrogel composites for osteochondral tissue regeneration. Appl Mater Today [Internet]. 2018;12:309-321. Available from: https://doi.org/10.1016/j.apmt.2018.06.005
- [70] Altieri MA, Nicholls CI. Agroecology Scaling Up for Food Sovereignty and Resiliency. Lichtfouse E (eds). Sustainable Agriculture Reviews [Internet]. Dordrecht: Springer Dordrecht; 2012. 1-29 p. Available from: https://doi.org/10.1007/978-94-007-5449-2
- [71] Ahmed S, Ikram S. Chitosan: Derivatives, Composites and Applications. Hoboken: Wiley; 2017. 516p.
- [72] Thomas MS, Koshy RR, Mary SK, Thomas S, et al. Starch, Chitin and Chitosan Based Composites and Nanocomposites [Internet]. Springer Cham; 2019. 57p. Available from: https://doi.org/10.1007/978-3-030-03158-9
- [73] Shakir M, Jolly R, Khan AA, Ahmed SS, et al. Resol based chitosan/ nano-hydroxyapatite nanoensemble for effective bone tissue engineering. Carbohydr Polym [Internet]. 2018;179:317-327. Available from: http://dx.doi.org/10.1016/j.carbpol.2017.09.103
- [74] Shen J, Jin B, Qi Y-C, Jiang Q, et al. Carboxylated chitosan/silverhydroxyapatite hybrid microspheres with improved antibacterial activity and cytocompatibility. Mater Sci Eng C [Internet]. 2017;78:589-597. Available from: http://dx.doi.org/10.1016/j.msec.2017.03.100
- [75] Costa-Pinto AR, Lemos AL, Tavaria FK, Pintado M. Chitosan and Hydroxyapatite Based Biomaterials to Circumvent Periprosthetic Joint Infections. Materials [Internet]. 2021;14(4):804. Available from: https://doi.org/10.3390/ma14040804
- [76] Li L, Iqbal J, Zhu Y, Zhang P, et al. Chitosan/Ag-hydroxyapatite nanocomposite beads as a potential adsorbent for the efficient removal of toxic aquatic pollutants. Int J Biol Macromol [Internet]. 2018;120:1752-1759. Available from: https://doi.org/10.1016/j.ijbiomac.2018.09.190
- [77] Nabipour H, Wang X, Song L, Hu Y. A fully bio-based coating made from alginate, chitosan and hydroxyapatite for protecting flexible polyurethane foam from fire. Carbohydr Polym [Internet]. 2020;246:116641. Available from: https://doi.org/10.1016/j.carbpol.2020.116641
- [78] Alvarez-Barreto J, Máquez K, Gallardo E, Moret J, et al. Mesenchymal Stem Cell Culture on Composite Hydrogels of Hydroxyapatite Nanoparticles and Photo-Crosslinking Chitosan. Rev Mex Ing Biom [Internet]. 2017;38(3):524-536. Available from: https://doi.org/10.17488/RMIB.38.3.2
- [79] Trakoolwannachai V, Kheolamai P, Ummartyotin S. Development of hydroxyapatite from eggshell waste and a chitosan-based composite: In vitro behavior of human osteoblast-like cell (Saos-2) cultures. Int J Biol Macromol [Internet]. 2019;134:557-564. Available from: https://doi.org/10.1016/j.ijbiomac.2019.05.004

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Juanita Daria Flores Valdez et al. Hydroxyapatite and Biopolymer Composites with Promising Biomedical Applications

- [80] Tripathi A, Saravanan S, Pattnaik S, Moorthi A, et al. Bio-composite scaffolds containing chitosan/nano-hydroxyapatite/nano-copper-zinc for bone tissue engineering. Int J Biol Macromol [Internet]. 2012;50(1):294-299. Available from: http://dx.doi.org/10.1016/j.ijbiomac.2011.11.013
- [81] Soriente A, Fasolino I, Gomez-Sánchez A, Prokhorov E, et al. Chitosan/hydroxyapatite nanocomposite scaffolds to modulate osteogenic and inflammatory response. J Biomed Mater Res Part A [Internet]. 2022;110(2):266-272. Available from: https://doi.org/10.1002/jbm.a.37283
- [82] Kaczmarek B, Sionkowska A, Skopinska-Wisniewska J. Influence of glycosaminoglycans on the properties of thin films based on chitosan/collagen blends. J Mech Behav Biomed Mater [Internet]. 2018;80:189-193. Available from: https://doi.org/10.1016/j.jmbbm.2018.02.006
- [83] Okada T, Nobunaga Y, Konishi T, Yoshioka T, et al. Preparation of chitosan-hydroxyapatite composite mono-fiber using coagulation method and their mechanical properties. Carbohydr Polym [Internet]. 2017;175:355-360. Available from: http://dx.doi.org/10.1016/j.carbpol.2017.07.072
- [84] Balagangadharan K, Chandran SV, Arumugam B, Saravanan S, et al. Chitosan/nano-hydroxyapatite/nano-zirconium dioxide scaffolds with miR-590-5p for bone regeneration. Int J Biol Macromol [Internet]. 2018;111:953-958. Available from: https://doi.org/10.1016/j.ijbiomac.2018.01.122
- [85] Chakravarty J, Rabbi MF, Chalivendra V, Ferreira T, et al. Mechanical and biological properties of chitin/polylactide (PLA)/ hydroxyapatite (HAP) composites cast using ionic liquid solutions. Int J Biol Macromol [Internet]. 2020;151:1213-1223. Available from: https://doi.org/10.1016/j.ijbiomac.2019.10.168
- [86] Akindoyo JO, Beg MDH, Ghazali S, Heim HP, et al. Effects of surface modification on dispersion, mechanical, thermal and dynamic mechanical properties of injection molded PLA-hydroxyapatite composites. Compos Part A Appl Sci Manuf [Internet]. 2017;103:96-105. Available from: http://dx.doi.org/10.1016/j.compositesa.2017.09.013
- [87] Grémare A, Guduric V, Bareille R, Heroguez V, et al. Characterization of printed PLA scaffolds for bone tissue engineering. J Biomed Mater Res Part A [Internet]. 2018;106(4):887-894. Available from: https://doi.org/10.1002/jbm.a.36289
- [88] Akindoyo JO, Beg MDH, Ghazali S, Heim HP, et al. Impact modified PLA-hydroxyapatite composites - Thermo-mechanical properties. Compos Part A Appl Sci Manuf [Internet]. 2018;107:326-333. Available from: https://doi.org/10.1016/j.compositesa.2018.01.017
- [89] Armentano I, Bitinis N, Fortunati E, Mattioli S, et al. Multifunctional nanostructured PLA materials for packaging and tissue engineering. Prog Polym Sci [Internet]. 2013;38(10-11):1720-1747. Available from: http://dx.doi.org/10.1016/j.progpolymsci.2013.05.010
- [90] Mohammadi MS, Bureau MN, Nazhat SN. Polylactic acid (PLA) biomedical foams for tissue engineering. In: Netti PA (eds). Biomedical Foams for Tissue Engineering Applications [Internet]. United Kingdom: Elsevier; 2014. 313-334. Available from: https://doi.org/10.1533/9780857097033.2.313
- [91] Esposito Corcione C, Gervaso F, Scalera F, Padmanabhan SK, et al. Highly loaded hydroxyapatite microsphere/ PLA porous scaffolds obtained by fused deposition modelling. Ceram Int [Internet]. 2019;45(2):2803-2810. Available from: https://doi.org/10.1016/j.ceramint.2018.07.297

- [92] Mondal S, Nguyen TP, Hiệp PV, Hoang G, et al. Hydroxyapatite nano bioceramics optimized 3D printed poly lactic acid scaffold for bone tissue engineering application. Ceram Int [Internet].
 2020;46(3):3443-3455. Available from: https://doi.org/10.1016/j.ceramint.2019.10.057
- [93] Talal A, Waheed N, Al-Masri M, McKay IJ, et al. Absorption and release of protein from hydroxyapatite-polylactic acid (HA-PLA) membranes. J Dent [Internet]. 2009;37(11):820-826. Available from: https://doi.org/10.1016/j.jdent.2009.06.014
- [94] Thanh DTM, Trang PTT, Thom NT, Phuong NT, et al. Effects of Porogen on Structure and Properties of Poly Lactic Acid/ Hydroxyapatite Nanocomposites (PLA/HAp). J Nanosci Nanotechnol [Internet]. 2016;16(9):9450-9459. Available from: https://doi.org/10.1166/jnn.2016.12032
- [95] Zhang H, Fu Q-W, Sun T-W, Chen F, et al. Amorphous calcium phosphate, hydroxyapatite and poly(D,L-lactic acid) composite nanofibers: Electrospinning preparation, mineralization and in vivo bone defect repair. Colloids Surf B [Internet]. 2015;136:27-36. Available from: http://dx.doi.org/10.1016/j.colsurfb.2015.08.015
- [96] Moura NKd, Siqueira IAWB, Machado JPdB, Kido HW, et al. Production and Characterization of Porous Polymeric Membranes of PLA/PCL Blends with the Addition of Hydroxyapatite. J Compos Sci [Internet]. 2019;3(2):45. Available from: https://doi.org/10.3390/jcs3020045
- [97] Zhang J, Wang Q, Wang A. In situ generation of sodium alginate/ hydroxyapatite nanocomposite beads as drug-controlled release matrices. Acta Biomater [Internet]. 2010;6(2):445-454. Available from: http://dx.doi.org/10.1016/j.actbio.2009.07.001
- [98] Sukhodub LF, Sukhodub LB, Litsis O, Prylutskyy Y. Synthesis and characterization of hydroxyapatite-alginate nanostructured composites for the controlled drug release. Mater Chem Phys [Internet]. 2018;217:228-234. Available from: https://doi.org/10.1016/j.matchemphys.2018.06.071
- [99] Rossi AL, Barreto IC, Maciel WQ, Rosa FP, et al. Ultrastructure of regenerated bone mineral surrounding hydroxyapatite-alginate composite and sintered hydroxyapatite. Bone [Internet]. 2012;50(1):301-310. Available from: http://dx.doi.org/10.1016/j.bone.2011.10.022
- [100] Rajkumar M, Meenakshisundaram N, Rajendran V. Development of nanocomposites based on hydroxyapatite/sodium alginate: Synthesis and characterisation. Mater Charact [Internet]. 2011;62(5):469-479. Available from: http://dx.doi.org/10.1016/j.matchar.2011.02.008
- [101] Rajesh R, Ravichandran YD. Development of a new carbon nanotube- alginate-hydroxyapatite tricomponent composite scaffold for application in bone tissue engineering. Int J Nanomedicine [Internet]. 2015;10:7-15. Available from: https://doi.org/10.2147/IJN.S79971
- [102] Mahmoud EM, Sayed M, El-Kady AM, Elsayed H, et al. In vitro and in vivo study of naturally derived alginate / hydroxyapatite bio composite scaffolds. Int J Biol Macromol [Internet]. 2020;165:1346-1360. Available from: https://doi.org/10.1016/j.ijbiomac.2020.10.014
- [103] Gholizadeh BS, Buazar F, Hosseini SM, Mousavi SM. Enhanced antibacterial activity, mechanical and physical properties of alginate/hydroxyapatite bionanocomposite film. Int J Biol Macromol [Internet]. 2018;116:786-792. Available from: https://doi.org/10.1016/j.ijbiomac.2018.05.104

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- [104] Bian T, Xing H. A collagen (Col)/nano-hydroxyapatite (nHA) biological composite bone scaffold with double multi-level interface reinforcement. Arab J Chem [Internet]. 2022;15(5):103733. Available from: https://doi.org/10.1016/j.arabjc.2022.103733
- [105] Bian T, Zhao K, Meng Q, Tang Y, et al. The construction and performance of multi-level hierarchical hydroxyapatite (HA)/ collagen composite implant based on biomimetic bone Haversian motif. Mater Des [Internet]. 2019;162:60-69. Available from: https://doi.org/10.1016/j.matdes.2018.11.040
- [106] Yilmaz E, Çakıroğlu B, Gökçe A, Findik F, et al. Novel hydroxyapatite/graphene oxide/collagen bioactive composite coating on Ti16Nb alloys by electrodeposition. Mater Sci Eng C [Internet]. 2019;101:292-305. Available from: https://doi.org/10.1016/j.msec.2019.03.078
- [107] Sun R-X, Lv Y, Niu Y-R, Zhao X-H, et al. Physicochemical and biological properties of bovine-derived porous hydroxyapatite/ collagen composite and its hydroxyapatite powders. Ceram Int [Internet]. 2017;43(18):16792-16798. Available from: http://dx.doi.org/10.1016/j.ceramint.2017.09.075
- [108] Li H, Sun X, Li Y, Wang H, et al. Carbon nanotube-collagen@ hydroxyapatite composites with improved mechanical and biological properties fabricated by a multi in situ synthesis process. Biomed Microdevices [Internet]. 2020;22:64. Available from: https://doi.org/10.1007/s10544-020-00520-5
- [109] Siswanto S, Hikmawati D, Kulsum U, Rudyardjo DI, et al. Biocompatibility and osteoconductivity of scaffold porous composite collagen-hydroxyapatite based coral for bone regeneration. Open Chem [Internet]. 2020;18(1):584-590. Available from: https://doi.org/10.1515/chem-2020-0080
- [110] Kane RJ, Weiss-Bilka HE, Meagher MJ, Liu Y, et al. Hydroxyapatite reinforced collagen scaffolds with improved architecture and mechanical properties. Acta Biomater [Internet]. 2015;17:16-25. Available from: http://dx.doi.org/10.1016/j.actbio.2015.01.031
- [111] Sionkowska A, Kozłowska J. Properties and modification of porous 3-D collagen/hydroxyapatite composites. Int J Biol Macromol [Internet]. 2013;52(1):250-259. Available from: http://dx.doi.org/10.1016/j.ijbiomac.2012.10.002
- [112] Zvicer J, Medic A, Veljovic D, Jevtic S, et al. Biomimetic characterization reveals enhancement of hydroxyapatite formation by fluid flow in gellan gum and bioactive glass composite scaffolds. Polym Test [Internet]. 2019;76:464-472. Available from: https://doi.org/10.1016/j.polymertesting.2019.04.004
- [113] Bastos AR, Raquel Maia F, Miguel Oliveira J, Reis RL, et al. Influence of gellan gum-hydroxyapatite spongy-like hydrogels on human osteoblasts under long-term osteogenic differentiation conditions. Mater Sci Eng C [Internet]. 2021;129:112413. Available from: https://doi.org/10.1016/j.msec.2021.112413
- [114] Anandan D, Madhumathi G, Nambiraj NA, Jaiswal AK. Gum based 3D composite scaffolds for bone tissue engineering applications. Carbohydr Polym [Internet]. 2019;214:62-70. Available from: https://doi.org/10.1016/j.carbpol.2019.03.020
- [115] Jamshidi P, Chouhan G, Williams RL, Cox SC, et al. Modification of gellan gum with nanocrystalline hydroxyapatite facilitates cell expansion and spontaneous osteogenesis. Biotechnol Bioeng [Internet]. 2016;113(7):1568-1576. Available from: https://doi.org/10.1002/bit.25915

- [116] Manda MG, da Silva LP, Cerqueira MT, Pereira DR, et al. Gellan gum-hydroxyapatite composite spongy-like hydrogels for bone tissue engineering. J Biomed Mater Res Part A [Internet]. 2018;106(2):479-490. Available from: https://doi.org/10.1002/jbm.a.36248
- [117] dos Santos MVB, Bastos Nogueira Rocha L, Gomes Vieira E, Leite Oliveira A, et al. Development of Composite Scaffolds Based on Cerium Doped-Hydroxyapatite and Natural Gums–Biological and Mechanical Properties. Materials [Internet]. 2019;12(15):2389. Available from: https://doi.org/10.3390/ma12152389
- [118] Heidari F, Razavi M, E.Bahrololoom M, Bazargan-Lari R, et al. Mechanical properties of natural chitosan/hydroxyapatite/ magnetite nanocomposites for tissue engineering applications. Mater Sci Eng C [Internet]. 2016;65:338-344. Available from: http://dx.doi.org/10.1016/j.msec.2016.04.039
- [119] Shavandi A, Bekhit AE-DA, Ali MA, Sun Z, et al. Development and characterization of hydroxyapatite/β-TCP/chitosan composites for tissue engineering applications. Mater Sci Eng C [Internet]. 2015;56:481-493. Available from: http://dx.doi.org/10.1016/j.msec.2015.07.004
- [120] Ghosh S, Ghosh S, Pramanik N. Bio-evaluation of doxorubicin (DOX)-incorporated hydroxyapatite (HAp)-chitosan (CS) nanocomposite triggered on osteosarcoma cells. Adv Compos Hybrid Mater [Internet]. 2020;3(3):303-314. Available from: https://doi.org/10.1007/s42114-020-00154-4
- [121] Gritsch L, Maqbool M, Mouriño V, Ciraldo FE, et al. Chitosan/ hydroxyapatite composite bone tissue engineering scaffolds with dual and decoupled therapeutic ion delivery: copper and strontium. J Mater Chem B [Internet]. 2019;7(40):6109-6124. Available from: https://doi.org/10.1039/C9TB00897G
- [122] Chen L, Hu J, Shen X, Tong H. Synthesis and characterization of chitosan-multiwalled carbon nanotubes/hydroxyapatite nanocomposites for bone tissue engineering. J Mater Sci: Mater Med [Internet]. 2013;24(8):1843-1851. Available from: https://doi.org/10.1007/s10856-013-4954-x
- [123] Li X, Nan K, Shi S, Chen H. Preparation and characterization of nano-hydroxyapatite/chitosan cross-linking composite membrane intended for tissue engineering. Int J Biol Macromol [Internet]. 2012;50(1):43-49. Available from: http://dx.doi.org/10.1016/j.ijbiomac.2011.09.021
- [124] Saravanan S, Nethala S, Pattnaik S, Tripathi A, et al. Preparation, characterization and antimicrobial activity of a bio-composite scaffold containing chitosan/nano-hydroxyapatite/nano-silver for bone tissue engineering. Int J Biol Macromol [Internet]. 2011;49(2):188-193. Available from: http://dx.doi.org/10.1016/j.ijbiomac.2011.04.010
- [125] Zhang J, Nie J, Zhang Q, Li Y, et al. Preparation and characterization of bionic bone structure chitosan/hydroxyapatite scaffold for bone tissue engineering. J Biomater Sci Polym Ed [Internet]. 2014;25(1):61-74. Available from: https://doi.org/10.1080/09205063.2013.836950
- [126] Carfi Pavia F, Conoscenti G, Greco S, La Carrubba V, et al. Preparation, characterization and in vitro test of composites polylactic acid/hydroxyapatite scaffolds for bone tissue engineering. Int J Biol Macromol [Internet]. 2018;119:945-953. Available from: https://doi.org/10.1016/j.ijbiomac.2018.08.007

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- [127] Ma B, Han J, Zhang S, Liu F, et al. Hydroxyapatite nanobelt/ polylactic acid Janus membrane with osteoinduction/barrier dual functions for precise bone defect repair. Acta Biomater [Internet]. 2018;71:108-117. Available from: https://doi.org/10.1016/j.actbio.2018.02.033
- [128] Anita Lett J, Sagadevan S, Paiman S, Mohammad F, et al. Exploring the thumbprints of Ag-hydroxyapatite composite as a surface coating bone material for the implants. J Mater Res Technol [Internet]. 2020;9(6):12824-12833. Available from: https://doi.org/10.1016/j.jmrt.2020.09.037
- [129] Chuan D, Fan R, Wang Y, Ren Y, et al. Stereocomplex poly(lactic acid)-based composite nanofiber membranes with highly dispersed hydroxyapatite for potential bone tissue engineering. Compos Sci Technol [Internet]. 2020;192:108107. Available from: https://doi.org/10.1016/j.compscitech.2020.108107
- [130] Zare RN, Doustkhah E, Assadi MHN. Three-dimensional bone printing using hydroxyapatite-PLA composite. Mater Today Proc [Internet]. 2021;42(3):1531-1533. Available from: https://doi.org/10.1016/j.matpr.2019.12.046
- [131] Alksne M, Kalvaityte M, Simoliunas E, Rinkunaite I, et al. In vitro comparison of 3D printed polylactic acid/hydroxyapatite and polylactic acid/bioglass composite scaffolds: Insights into materials for bone regeneration. J Mech Behav Biomed Mater [Internet]. 2020;104:103641. Available from: https://doi.org/10.1016/j.jmbbm.2020.103641
- [132] Zhang H, Mao X, Zhao D, Jiang W, et al. Three dimensional printed polylactic acid-hydroxyapatite composite scaffolds for prefabricating vascularized tissue engineered bone: An in vivo bioreactor model. Sci Rep [Internet]. 2017;7(1):15255. Available from: http://dx.doi.org/10.1038/s41598-017-14923-7